

Determination of Dissolved Inorganic Ammonium (NH₄) in Fresh/Estuarine/Coastal Waters

1. SCOPE and APPLICATION

- 1.1 Determination of ammonium is by the Bertholet Reaction in which a blue-colored compound, similar to indophenol, forms when a solution of ammonium salt is added to sodium phenoxide. The method is used to analyze all ranges of salinity.
- 1.2 A Method Detection Limit (MDL) of 0.001 mg NH₄-N/L was determined using the Student's *t* value (3.14) times the standard deviation of seven low level replicates. If more than seven replicates are used to determine the MDL, refer to the Student's *t* test table for the appropriate n-1 value.
- 1.3 The Quantitation Limit for NH₄ was set at 0.010 mg NH₄-N/L.
- 1.4 The method is suitable for NH₄ concentrations 0.001 to 1.68 mg NH₄-N/L.
- 1.5 This procedure should be used by analysts experienced in the theory and application of aqueous inorganic analysis. Three months experience with an analyst, experienced in the analysis of ammonium in aqueous samples, is required.
- 1.6 This method can be used for all programs that require analysis of dissolved ammonium.
- 1.7 This procedure conforms to EPA Method 350.2 (1979).

2. SUMMARY

2.1 Filtered samples are complexed with sodium potassium tartrate and sodium citrate. The complexed sample reacts with alkaline phenol and hypochlorite, catalyzed by sodium nitroprusside, yielding an intense blue color suitable for photometric measurement.

3. DEFINITIONS

- 3.1 Acceptance Criteria – Specified limits placed on characteristics of an item, process, or service defined in a requirement document. (ASQC)
- 3.2 Accuracy – The degree of agreement between an observed value and an accepted reference value. Accuracy includes a combination of random error (precision) and systematic error (bias) components which are due to sampling and analytical operations; a data quality indicator. (QAMS)
- 3.3 Aliquot – A discrete, measured, representative portion of a sample taken for analysis. (EPA QAD Glossary)
- 3.4 Analytical Range – 0.010 to 1.68 mg NH₄-N/L. The overall analytical range is comprised of two distinct concentration ranges. A separate

calibration is performed for each range. These ranges include 0.010 to 0.168 mg NH₄-N/L, and 0.168 to 1.68 mg NH₄-N/L. Two sub-ranges are utilized so that samples can be analyzed on the most appropriate scale possible.

- 3.5 Batch – Environmental samples, which are prepared and /or analyzed together with the same process and personnel, using the same lot(s) of reagents. A **preparation batch** is composed of one to 200 environmental samples of the same matrix, meeting the above mentioned criteria and with a maximum time between the start of processing of the first and last sample in the batch to be 8 hours. An **analytical batch** is composed of prepared environmental samples (extracts, digestates, concentrates) and/or those samples not requiring preparation, which are analyzed together as a group using the same calibration curve or factor. An analytical batch can include samples originating from various environmental matrices and can exceed 20 samples. (NELAC/EPA)
- 3.6 Blank- A sample that has not been exposed to the analyzed sample stream in order to monitor contamination during sampling, transport, storage or analysis. The blank is subjected to the usual analytical and measurement process to establish a zero baseline or background value and is sometimes used to adjust or correct routine analytical results. (ASQC)
- 3.7 Calibrate- To determine, by measurement or comparison with a standard, the correct value of each scale reading on a device. The levels of the applied calibration standard should bracket the range of planned or expected sample measurements. (NELAC)
- 3.8 Calibration – The set of operations which establish, under specified conditions, the relationship between values indicated by a measuring device. The levels of the applied calibration standard should bracket the range of planned or expected sample measurements. (NELAC)
- 3.9 Calibration Blank – A volume of reagent water fortified with the same matrix as the calibration standards, without analyte added.
- 3.10 Calibration Curve – The graphical relationship between known values, such as concentrations, or a series of calibration standards and their analytical response. (NELAC)
- 3.11 Calibration Method – A defined technical procedure for performing a calibration. (NELAC)
- 3.12 Calibration Standard – A substance or reference material used to calibrate an instrument. (QAMS)
 - 3.12.1 Initial Calibration Standard (STD) – A series of standard solutions used to initially establish instrument calibration responses and develop calibration curves for individual target analytes.
 - 3.12.2 Initial Calibration Verification (ICV) – An individual standard, distinct from the Initial Calibration Standards (STD), analyzed initially, prior to any sample analysis,

which verifies acceptability of the calibration curve or previously established calibration curve.

- 3.12.3 Continuing Calibration Verification (CCV) – An individual standard, distinct from the Initial Calibration Standards (STD), analyzed after every 15-20 field sample analyses.
- 3.13 Certified Reference Material (CRM) – A reference material one or more of whose property values are certified by a technically valid procedure, accompanied by or traceable to a certificate or other documentation which is issued by a certifying body. (ISO 17025)
- 3.14 Corrective Action – Action taken to eliminate the causes of an existing nonconformity, defect or other undesirable situation in order to prevent recurrence. (ISO 8402)
- 3.15 Deficiency – An unauthorized deviation from acceptable procedures or practices. (ASQC)
- 3.16 Demonstration of Capability – A procedure to establish the ability of the analyst to generate acceptable accuracy. (NELAC)
- 3.17 Detection Limit – The lowest concentration or amount of the target analyte that can be determined to be different from zero by a single measurement at a stated degree of confidence.
- 3.18 Duplicate Analysis – The analyses of measurements of the variable of interest performed identically on two sub samples (aliquots) of the same sample. The results from duplicate analyses are used to evaluate analytical or measurement precision but not the precision of sampling, preservation or storage internal to the laboratory. (EPA-QAD)
- 3.19 External Standard (ES) – A pure analyte (Ammonium Sulfate $(\text{NH}_4)_2\text{SO}_4$) that is measured in an experiment separate from the experiment used to measure the analyte(s) in the sample. The signal observed for a known quantity of the pure external standard is used to calibrate the instrument response for the corresponding analyte(s). The instrument response is used to calculate the concentrations of the analyte(s) in the unknown sample.
- 3.20 Field Duplicates (FD1 and FD2) – Two separate samples collected at the same time and place under identical circumstances and treated exactly the same throughout field and laboratory procedures. Analyses of FD1 and FD2 provide a measure of the precision associated with sample collection, preservation and storage, as well as with laboratory procedures.
- 3.21 Holding time – The maximum time that samples may be held prior to analysis and still be considered valid. (40 CFR Part 136) The time elapsed from the time of sampling to the time of extraction or analysis, as appropriate.
- 3.22 Laboratory Duplicates (LD1 and LD2) – Two aliquots of the same sample taken in the laboratory and analyzed separately with identical procedures. Analyses of LD1 and LD2 indicate precision associated with laboratory procedures, but not with sample collection, preservation, or storage procedures.

- 3.23 Laboratory Reagent Blank (LRB) – A blank matrix (i.e., DI water) that is treated exactly as a sample including exposure to all glassware, equipment, solvents, and reagents that are used with other samples. The LRB is used to determine if method analytes or other interferences are present in the laboratory environment, the reagents, or the instrument.
- 3.24 Laboratory Control Sample (LCS) – A sample matrix, free from the analytes of interest, spiked with verified known amounts of analytes from a source independent of the calibration standard or a material containing known and verified amounts of analytes. The LCS is generally used to establish intra-laboratory or analyst-specific precision and bias or to assess the performance of all or a portion of the measurement system. (NELAC)
- 3.25 Limit of Detection (LOD) – The lowest concentration level that can be determined by a single analysis and with a defined level of confidence to be statistically different from a blank. (ACS)
- 3.26 Limit of Quantitation (LOQ) – The minimum levels, concentrations, or quantities of a target variable (target analyte) that can be reported with a specified degree of confidence. The LOQ is set at 3 to 10 times the LOD, depending on the degree of confidence desired.
- 3.27 Linear Dynamic Range (LDR) – The absolute quantity over which the instrument response to an analyte is linear. This specification is also referred to as the Linear Calibration Range (LCR).
- 3.28 Material Safety Data Sheets (MSDS) – Written information provided by vendors concerning a chemical’s toxicity, health hazards, physical properties, fire, and reactivity data including storage, spill, and handling precautions.
- 3.29 May – Denotes permitted action, but not required action. (NELAC)
- 3.30 Method Detection Limit (MDL) – The minimum concentration of an analyte that can be identified, measured, and reported with 99% confidence that the analyte concentration is greater than zero (Standard Methods).
- 3.31 Must – Denotes a requirement that must be met. (Random House College Dictionary)
- 3.32 Photometer – measures the absorbance of the solution in the cell in a multicell cuvette. Light passes from the lamp through the condensing lenses to the interference filter. The plane surface of the first condensing lens is coated with a material which reflects heat and infrared light. The filters are mounted on a filter wheel. There are 15 positions for filters. Each filter corresponds to a wavelength of interest. The 630 nm filter is specified by the test definition for ammonium. After passing through the filter the light is converted into a stream of light pulses by a chopper. Then the light is directed via a quartz fiber through a focusing lens and a slit to the beam divider. The

beam divider divides the light into two parts. A specified portion is reflected to the reference detector, which monitors the light level fluctuations. The remaining major portion of the light beam goes through the liquid in the cell to the signal detector, which measures the amount of light absorbed.

- 3.33 Precision – The degree to which a set of observations or measurements of the same property, obtained under similar conditions, conform to themselves; a data quality indicator. Precision is usually expressed as standard deviation, variance or range, in either absolute or relative terms. (NELAC)
- 3.34 Preservation – Refrigeration, freezing, and/or reagents added at the time of sample collection (or later) to maintain the chemical and or biological integrity of the sample.
- 3.35 Quality Control Sample (QCS) – A sample of analyte of known and certified concentration. The QCS is obtained from a source external to the laboratory and different from the source of calibration standards. It is used to check laboratory performance with externally prepared test materials.
- 3.36 Run Cycle – Typically a day of operation – the entire analytical sequence from sampling the first standard to the last sample of the day.
- 3.37 Sample Segment – Bar-coded metal tray that holds up to fourteen four milliliter auto analyzer vials containing samples or standards. The user identifies each vial in the operating software.
- 3.38 Sample Segment Holder – An automated temperature controlled carousel that contains up to six sample segments. This carousel spins in clockwise or counterclockwise manner to move the sample segments into position for analysis. This carousel format allows for continuous processing.
- 3.39 Sensitivity – The capability of a test method or instrument to discriminate between measurement responses representing different levels (concentrations) of a variable of interest.
- 3.40 Shall – Denotes a requirement that is mandatory whenever the criterion for conformance with the specification requires that there be no deviation. (ANSI)
- 3.41 Should – Denotes a guideline or recommendation whenever noncompliance with the specification is permissible. (ANSI)
- 3.42 Standard Reference Material (SRM) – Material which has been certified for specific analytes by a variety of analytical techniques and/or by numerous laboratories using similar analytical techniques. These may consist of pure chemicals, buffers, or compositional standards. The materials are used as an indication of the accuracy of a specific analytical technique.
- 3.43 Test Definition – A photometric test consisting of a user defined testing sequence, reagent additions, calibration standards, incubations and absorption results.

3.44 Test Flow – Functions to define the parameter for reagent and sample dispensing, dilution, incubation and measurement.

4 INTERFERENCES

- 4.1 Suspended matter in the sample will scatter light as it passes through the cuvette to the detector. High blank responses will result. The identified sample will be reanalyzed.
- 4.2 Blemishes in the cuvette, as result of the manufacturing process, will result in high blank responses. The identified sample will be reanalyzed.
- 4.3 Seawater contains calcium and magnesium ions in sufficient concentrations to cause precipitation during analysis, reducing color production. Adding sodium potassium tartrate and sodium citrate reduces the interference.
- 4.4 Eliminate any added acidity in samples because intensity of measured color is pH-dependent.

5 SAFETY

- 5.1 Safety precautions must be taken when handling reagents, samples and equipment in the laboratory. Protective clothing including lab coats, safety glasses and enclosed shoes should be worn. In certain situations, it will be necessary to also use gloves and/or a face shield. If solutions come in contact with eyes, flush with water continuously for 15 minutes. If solutions come in contact with skin, wash thoroughly with soap and water. Contact Solomons Rescue Squad (911) if emergency treatment is needed and also inform the CBL Business Manager of the incident. Contact the CBL Business Manager if additional treatment is required.
- 5.2 The toxicity or carcinogenicity of each reagent used in this procedure may not have been fully established. Each chemical should be regarded as a potential health hazard and exposure should be as low as reasonably achievable. Cautions are included for known hazardous materials and procedures.
- 5.3 Do not wear jewelry when troubleshooting electrical components. Even low voltage points are dangerous and can injure if allowed to short circuit.
- 5.4 The following hazard classifications are listed for the chemicals used in this procedure. Detailed information is provided on Material Safety Data Sheets (MSDS).

Chemical	Health	Flammability	Reactivity	Contact	Storage
Sodium potassium tartrate	0	1	0	0	Green
Sodium citrate	1	1	0	1	Green
Sulfuric acid	4	0	2	4	White
Phenol	3	2	1	4	White Stripe
Sodium hydroxide	3	0	2	4	White Stripe

Sodium nitroprusside	3	0	0	1	Blue
Sodium hypochlorite (Clorox)	3	0	2	4	White
Ammonium sulfate	1	0	1	2	Green
Chloroform	3	1	1	3	Blue

On a scale of 0 to 4 the substance is rated on four hazard categories: health, flammability, reactivity, and contact. (0 is non-hazardous and 4 is extremely hazardous)

STORAGE

Red – Flammability Hazard: Store in a flammable liquid storage area.

Blue – Health Hazard: Store in a secure poison area.

Yellow – Reactivity Hazard: Keep separate from flammable and combustible materials.

White – Contact Hazard: Store in a corrosion-proof area.

Green – Use general chemical storage (On older labels, this category was orange).

Striped – Incompatible materials of the same color class have striped labels. These products should not be stored adjacent to substances with the same color label. Proper storage must be individually determined.

6 EQUIPMENT AND SUPPLIES

6.1 Aquakem 250 multi-wavelength automated discrete photometric analyzer.

Aquakem 250 control software operates on a computer running Microsoft Windows NT or XP operating system.

6.2 Freezer, capable of maintaining $-20 \pm 5^\circ \text{C}$.

6.3 Lab ware – All reusable lab ware (glass, Teflon, plastic, etc) should be sufficiently clean for the task objectives. This laboratory cleans all lab ware related to this method with a 10% HCl (v/v) acid rinse.

7 REAGENTS AND STANDARDS

7.1 Purity of Water – Unless otherwise indicated, references to water shall be understood to mean reagent water conforming to Specification D 1193, Type I. Freshly prepared water should be used for making the standards intended for calibration. The detection limits of this method will be limited by the purity of the water and reagents used to make the standards.

7.2 Purity of Reagents – Reagent grade chemicals shall be used in all tests. Unless otherwise indicated, it is intended that all reagents shall conform to specifications of the Committee on Analytical Reagents of the American Chemical Society, where such specifications are available. Other grades may be used, provided it is first ascertained that the reagent is of sufficiently high purity to permit its use without compromising the accuracy of the determination.

7.3 Complexing reagent

Sodium potassium tartrate ($\text{KNaC}_4\text{H}_4\text{O}_6 \cdot 4\text{H}_2\text{O}$)	19.5 g
Sodium citrate ($\text{C}_6\text{H}_8\text{N}_2 \text{O}_2\text{S}$)	14.0 g
Sulfuric acid (H_2SO_4), concentrated (sp. gr. 1.84)	as required

In a 1000 mL beaker, dissolve 19.5 g sodium potassium tartrate and 14.0 g sodium citrate in approximately 500 mL deionized water. Place on heated magnetic stir plate. After dissolution, add 1 mL 5M NaOH and allow to boil uncovered until volume is reduced to 300 mL. Remove from heat, cool and add 1 mL 5N HCl. Bring volume to ~400 mL with deionized water and adjust the pH of the solution to 5.0 using concentrated sulfuric acid. Transfer solution to a 500 mL volumetric flask and bring to volume with deionized water. Write name of preparer, preparation date, reagent manufacturer, manufacturer's lot number in the Analytical Reagent log book. Store flask in dark at room temperature. Reagent is stable for six months.

7.4 Alkaline phenol solution

Phenol (C ₆ H ₅ OH), 88%	23.6 mL
Sodium hydroxide (NaOH), 50% (w/w)	18.0 g

In a 250 mL volumetric flask, slowly add 23.6 mL of 88% phenol to approximately 150 mL deionized water. While in an ice bath, add exactly 18 g of 50% (w/w) sodium hydroxide to the phenol/water solution. Bring flask to volume with deionized water. Store reagent in light resistant container in refrigerator. Write name of preparer, preparation date, reagent manufacturer, manufacturer's lot number in the Analytical Reagent log book. Reagent is stable for two weeks.

WARNING: Phenol is extremely dangerous and should be handled accordingly. Prepare in an operating fume hood wearing protective gloves.

7.5 Sodium hypochlorite, 1% –

Sodium hypochlorite (Clorox Regular – 8.25%)	32 mL
Deionized water	200 mL

In a 250 mL brown poly bottle, dilute 32 mL of commercially available bleach containing 8.25% sodium hypochlorite with 200 mL deionized water. Write name of preparer, preparation date, manufacturer, date Clorox purchased in the Analytical Reagent log book. Reagent is stable for two days at room temperature.

7.6 Sodium nitroprusside (Sodium nitroferricyanide), 0.05% –

Sodium nitroprusside	0.5 g
Deionized water	up to 1000 mL

In a 1000 mL flask, dissolve 0.5 g of sodium nitroprusside in 900 mL deionized water. Dilute to 1000 mL with deionized water and transfer to brown poly bottle. Write name of preparer, preparation date, reagent manufacturer, manufacturer lot number in the Analytical Reagent log book. Reagent is stable for 12 months at room temperature.

7.7 Ammonium Stock Standard, 1,500 μM –

Ammonium Sulfate [(NH ₄) ₂ SO ₄], primary standard grade, dried at 45°C	0.100 g
Deionized water	up to 1000 mL

In a 1000 mL volumetric flask, dissolve 0.100 g of ammonium sulfate in ~800 mL of deionized water. Dilute to 1000 mL with deionized water (1 mL contains 1.5 μ moles N). Transfer to glass bottle. Add 1 mL of chloroform as a preservative. Write name of preparer, preparation date, standard manufacturer, manufacturer lot number in the Analytical Standard log book. Store at room temperature. Make fresh every 4-6 months.

7.8 Working Regular Ammonium Standard –

Stock Ammonium Standard	0.80 mL
Deionized water	up to 100 mL

In a volumetric flask, dilute 0.80 mL of Stock Ammonium Standard to 100 mL with deionized water to yield a concentration of 12 μ M NH_4 –N/L (0.168 mg N/L). Write name of preparer, preparation date, Ammonium Stock Standard preparation date in the Analytical Standard log book. Make fresh every month.

7.9 Working High Ammonium Standard –

Stock Ammonium Standard	8.00 mL
Deionized water	up to 100 mL

In a volumetric flask, dilute 8.00 mL of Stock Ammonium Standard to 100 mL with deionized water to yield a concentration of 120 μ M NH_4 –N/L (1.68 mg N/L). Write name of preparer, preparation date, Ammonium Stock Standard preparation date in the Analytical Standard log book. Make fresh every month.

7.10 Aquakem Cleaning Solution –

Clorox	75.0 mL
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In a 100 mL volumetric flask, dilute 75.0 mL of Clorox to volume with deionized water to yield a concentration of 75% Clorox. Recent (2012) trends in commercially available Clorox, have necessitated altering this formula to 55.0 mL Clorox in 100 mL flask. Write name of preparer, preparation date, reagent manufacturer, manufacturer lot number in the Analytical Reagent log book. Reagent is stable for six months.

8 SAMPLE COLLECTION, PRESERVATION, AND STORAGE

8.1 Water collected for ammonium should be filtered through a Whatman GF/F glass fiber filter (nominal pore size 0.7 μ m), or equivalent.

8.2 Water collected for ammonium should be measured for salinity.

8.3 Water collected for ammonium should be frozen at -20° C. The AutoAnalyzer vial container should be clean and sample rinsed.

8.4 Frozen ammonium samples may be stored up to 28 days. It has been shown that frozen QCS samples up to a year old still fall well within the control limits.

8.5 Ammonium samples may be refrigerated at 4° C for no longer than one day.

9 QUALITY CONTROL

9.1 The laboratory is required to operate a formal quality control (QC) program. The minimum requirements of this program consist of an initial demonstration of laboratory capability and the continued analysis of laboratory instrument blanks and calibration standard material, analyzed as samples, as a continuing check on performance. The laboratory is required to maintain performance records that define the quality of data generated.

9.2 Initial Demonstration of Performance

9.2.1 The initial demonstration of capability (nitrite) – is used to characterize instrument performance (MDLs) and laboratory performance (analysis of QC samples) prior to the analyses conducted by this procedure.

9.2.2 Linear Dynamic Range – LDR (Linear Calibration Range) should be established for ammonium using appropriate five point calibration curve.

9.2.3 Quality Control Sample (QCS/SRM) – When using this procedure, a quality control sample is required to be analyzed at the beginning and end of the run, to verify data quality and acceptable instrument performance. If the determined concentrations are not within $\pm 10\%$ of the certified values, performance of the determinative step of the method is unacceptable. The source of the problem must be identified and corrected before either proceeding with the initial determination of MDLs or continuing with analyses.

9.2.4 Method Detection Limits (MDLs) – MDLs should be established for ammonium using a low level ambient water sample. To determine the MDL values, analyze seven replicate aliquots of water. Perform all calculations defined in the procedure (Section 12) and report the concentration values in the appropriate units. Calculate the MDL as follows: $MDL = St_{(n-1, 1-\alpha=0.99)}$

Where, S = Standard deviation of the replicate analyses.

n=number of replicates

$t_{(n-1, 1-\alpha=0.99)}$ = Student's *t* value for the 99%

confidence level with n-1 degrees of freedom

($t=3.14$ for 7 replicates.)

- 9.2.5 MDLs shall be determined yearly and whenever there is a significant change in instrument response, a significant change in instrument configuration, or a new matrix is encountered.

9.3 Assessing Laboratory Performance

- 9.3.1 Laboratory Reagent Blank (LRB) – The laboratory must analyze at least one LRB with each batch of samples. The LRB consists of Nanopure water treated the same as the samples. Analyte found in LRB indicates possible reagent or laboratory environment contamination. LRB data are used to assess and correct contamination from the laboratory environment. LRB above the lowest standard requires that the source of the problem must be identified and corrected before proceeding with analyses.
- 9.3.2 Quality Control Sample (QCS)/ Standard Reference Material (SRM) – When using this procedure, a quality control sample is required to be analyzed at the beginning of the run and end of the run, to verify data quality and acceptable instrument performance. If the determined concentrations are not within $\pm 3s$ of the certified values, performance of the determinative step of the method is unacceptable. The source of the problem must be identified and corrected before either proceeding with the initial determination of MDLs or continuing with the analyses. The results of these QCS/SRM samples shall be used to determine batch acceptance.
- 9.3.3 The QCS are obtained from a source external to the laboratory and different from the source of calibration standards.
- 9.3.4 Control Charts – The Accuracy Control Chart for QCS/SRM samples is constructed from the average and standard deviation of the 20 most recent QCS/SRM measurements. The accuracy chart includes upper and lower warning levels ($WL=\pm 2s$) and upper and lower control levels ($CL=\pm 3s$). These values are derived from stated values of the QCS/SRM. The standard deviation (s) is specified relative to statistical confidence levels of 95% for WLs and 99% for CLs. Set up an accuracy chart by using percent recovery since the concentration of the QCS/SRM varies. Enter QCS/SRM results on the chart each time the sample is analyzed
- 9.3.5 Continuing Calibration Verification (CCV) – Following every 18-23 samples, one CCV of 6.0 μM $\text{NH}_4\text{-N/L}$ (0.126 mg N/L) Regular NH_4 , 60 μM $\text{NH}_4\text{-N/L}$ (1.26 mg N/L) NH_4HIGH , is analyzed to assess instrument performance. The CCVs are made from the same material as calibration standards [$(\text{NH}_4)_2\text{SO}_4$], and are to be within $\text{TV} \pm 3s$. Failure to meet the criteria requires correcting the problem, including reanalysis of any affected samples. If not enough sample exists, the data must be qualified if reported.
- 9.3.6 Reagent Blank – The Reagent Blank Control Chart for Reagent Blank samples is constructed from the average and standard deviation of the

20 most recent Reagent Blank measurements. The accuracy chart includes upper and lower warning levels ($WL=\pm 2s$) and upper and lower control levels ($CL=\pm 3s$). The standard deviation (s) is specified relative to statistical confidence levels of 95% for WLs and 99% for CLs. Enter Reagent Blank results on the chart each time the Reagent Blank is analyzed.

9.4 Assessing Analyte Recovery - % Recovery

9.4.1 Analyte recovery is assessed through percent recoveries of laboratory spikes of samples.

9.4.2 $\% \text{ Recovery} = (\text{Actual value}/\text{Expected value}) \times 100$

9.5 Assessing Analyte Precision – Relative Percent Difference

9.5.1 Analyte replication is assessed through duplicate analyses of samples – Relative Percent Difference.

9.5.2 $RPD = (\text{Laboratory Duplicate Result 1} - \text{Laboratory Duplicate Result 2}) / [(\text{Laboratory Duplicate Result 1} + \text{Laboratory Duplicate Result 2}) / 2] \times 100$

9.6 Corrective Actions for Out of Control Data

9.6.1 Control limit – If one measurement exceeds Accuracy Control Chart CL, repeat the analysis immediately. If the repeat measurement is within the CL, continue analyses; if it exceeds the CL, discontinue analyses and correct the problem.

9.6.2 Warning limit – If two out of three successive points exceed Accuracy Control Chart WL, analyze another sample. If the next point is within WL, continue analyses; if the next point exceeds the WL, evaluate potential bias and correct the problem.

9.6.3 Trending – If seven successive Accuracy Control Chart measurements are on the same side of the central line, discontinue analyses and correct the problem.

9.6.4 When external QCS samples are out of control, correct the problem. Reanalyze the samples analyzed between the last in-control measurement and the out-of-control one.

9.6.5 When external CCV samples are out of control, correct the problem. Reanalyze the samples analyzed between the last in-control measurement and the out-of-control one.

9.7 General Operation - To assure optimal operation and analytical results, the Reagent Blank and CCV are tracked daily in the raw data file, copied to Reagent Blank and CCV Control Charts.

10 CALIBRATION AND STANDARDIZATION

10.1 Calibration – Daily calibration must be performed before sample analysis may begin. Five or six point calibrations are used with each of the two sub-

calibrations that cover the analytical range. Two working ammonium standards are used to produce the calibrators for each set of two calibration curves. The instrument performs serial dilutions of working standards to produce the five or six calibrators defined for each curve. The following outlines the preparation of the working standards and the following table describes the subsequent serial dilutions the instrument performs to make each standard for each of the two calibration curves.

Ammonium Working Standards:

Regular NH4 (NH4CBL2)

Working Standard 0.168 mg N/L (0.8 mL stock standard to 100 mL)

Working CCV 0.126 mg N/L (0.6 mL stock standard to 100 mL)

NH4HIGH

Working Standard 1.68 mg N/L (8.0 mL stock standard to 100 mL)

Working CCV 1.26 mg N/L (6.0 mL stock standard to 100 mL)

Nitrite Calibrators:

Test Name	Working Standard	Dilution Factor	Concentration mg N/L
NH4CBL2	0.168 mg N/L	1+9	0.0168
	0.168 mg N/L	1+4	0.0336
	0.168 mg N/L	1+2	0.056
	0.168 mg N/L	1+1	0.084
	0.168 mg N/L	1+0	0.168
NH4HIGH	1.68 mg N/L	1+9	0.168
	1.68 mg N/L	1+6	0.24
	1.68 mg N/L	1+3	0.42
	1.68 mg N/L	1+2	0.56
	1.68 mg N/L	1+1	0.84
	1.68 mg N/L	1+0	1.68

10.2 The instrument software prepares a standard curve for each set of calibrators. A graph plotting measured absorbance against standard concentration is presented for review and approval. If acceptance criteria are not met the entire curve can be reanalyzed or individual standards can be reanalyzed. One standard value (original or reanalyzed) for each and every standard is incorporated in the curve. The coefficient of determination (Person's r value) for the calibration curve as well as the calculated concentration of each calibrator is reviewed. The calculated value of each calibrator must be within ten percent of the expected value. The coefficient of determination (Person's r value) for the calibration curve must be greater than 0.980.

11 PROCEDURE – DAILY OPERATIONS QUALITY CONTROL

- 11.1 Turn on computer. Computer will automatically initiate Konelab software. Once software is running, turn on instrument and allow connection between instrument and computer to complete.
- 11.2 Discard any water remaining in the water reservoir from the previous analytical run. Fill the water reservoir with fresh deionized water.
- 11.3 Remove samples from freezer that will be analyzed that day. Allow samples to begin thawing. Begin daily bench sheet documentation.
- 11.4 Once water reservoir is full, “perform washes” – complete five wash cycles and then initiate “start-up” at main menu.
- 11.5 Gather working standards and reagents from refrigerator or dark cabinet during startup. Assess standards and reagents. Remake anything that has exceeded the time over which it is considered stable.
- 11.6 Once startup is complete, check that the instrument water blank of water from the reservoir has performed within acceptance limits. If any of the instrument functions are outside their predefined and software controlled limits, the user will be notified on the main menu page. User takes corrective action to return instrument functions to controlled limits.
- 11.7 Load reagents in specified position in reagent carousel and place in refrigerated reagent compartment.
- 11.8 Load working standards in a sample segment, identify the standards in their positions from the drop down menus at the individual segment positions, and load into instrument.
- 11.9 Select the methods to be calibrated. Two methods will be calibrated – NH4CBL2, and NH4HIGH are the method names to be selected in the software.
- 11.10 Begin calibration – See test flow below for stepwise instrument functions for the analysis of standards and samples.

Test Flow – Method of Analysis, Stepwise

- 100 µL sample to cuvette with mixing
- 55 µL Complexing Reagent to cuvette with mixing
- 33 µL Alkaline Phenol Reagent to cuvette with mixing
- Blank response measurement at 630 nm
- 26 µL Sodium Hypochlorite Reagent to cuvette with mixing
- 33 µL Sodium Nitroprusside Reagent to cuvette with mixing
- Incubation, 420 seconds, 37°C
- End point absorbance measurement, 630 nm
- Software processes absorbance value, blank response value and uses calibration curve to calculate analyte concentration (mg/L N as NH₄)
- User is notified if any measured values used to calculate final concentration are outside preset limits. If so, analyst has options to accept result, rerun the sample or rerun the sample diluted to a user or software specified factor.
- User is notified of each blank response value. Blank response >0.005 absorbance units indicates a scratched cuvette or turbid sample. If the blank response value exceeds 0.005 absorbance units, the analyst

specifies that the sample is reanalyzed. If the blank response value of the reanalyzed sample is <0.005 absorbance units, the reanalyzed result is accepted. If the same concentration and blank response value >0.005 absorbance units is again obtained, the results are accepted.

- 11.11 Organize samples, reagent blanks, check standards and all quality control samples while instrument performs calibrations.
- 11.12 As calibration curves are produced by the instrument, review them for acceptability. The instrument software prepares a standard curve for each set of calibrators. A graph plotting measured absorbance against standard concentration is presented for review and approval. If acceptance criteria are not met, either the entire curve shall be reanalyzed or individual standards shall be reanalyzed, depending on the violation. One standard value (original or reanalyzed) for each and every calibrator is incorporated in the curve.
- 11.13 Once calibration curves are accepted, samples are loaded into the sample segments and loaded into the instrument for analysis. After the Reagent Blank, the first sample analyzed should be an ICV (initial calibration verification) sample. There should be one ICV sample for each calibration curve, of a concentration close to the middle of each range. The following are the usual ICV samples for each curve: 0.126 mg N/L for NH4CBL2 and 1.26 mg N/L for NH4HIGH.
- 11.14 Samples are loaded into the segments and analyzed. CCV (Continuing Calibration Verification) samples (one for each of the two calibration ranges) follow every 18-23 samples. Standard Reference Material (SRM) samples, as well as Laboratory Reagent Blanks (LRB) are scattered throughout the analytical batch. Throughout the analytical batch, samples are chosen as laboratory duplicates and laboratory spikes to assess analyte precision and analyte recovery, respectively. The total number of duplicates and spikes performed will be equal or greater to ten percent of the total number of samples in the analytical batch.
- 11.15 As sample analysis is complete, results must be reviewed and accepted manually. If results fall outside acceptance limits, the sample should be reanalyzed. If sample result exceeds the highest standard of the calibration range it was run within, the samples can be automatically diluted by the instrument and reanalyzed. If the result is such that it will fall within a higher calibration range, it should be reanalyzed in that range. If the result is such that it will fall within a lower calibration range, it should be reanalyzed within that range.
- 11.16 User reviews salinity of samples. If any sample salinity concentration is greater than 20 ppt, the analyst reanalyzes the sample diluted to a user specified factor to obtain a salinity ≤ 10 ppt.
- 11.17 Upon completion of all analysis, results are saved to a daily report file. The file is named by the run date. The daily report file for analytical batch of January 1, 2005 would be named 010105. The file is converted to Microsoft Excel for data work up. Remaining samples are discarded.
- 11.18 All reagents are removed from the reagent chamber and returned to the refrigerator or dark cabinet. Reagents that have exceeded their stability period are discarded.

- 11.19 Aquakem Cleaning Solution is inserted into the instrument and shut down procedures are initiated. Daily files are cleared from the instrument software, the software is exited and the instrument is shut down. The computer is shut down.
- 11.20 The waste is flushed down the drain with copious amounts of tap water. The waste cuvette box is moved to the fume hood.

12 DATA ANALYSIS AND CALCULATIONS

- 12.1 Upon completion of all analysis, results are saved to a daily report file. The file is named by the run date. The daily report file for analytical batch of January 1, 2005 would be named 010105. The file is converted to Microsoft Excel for data work up. The instrument software has calculated final sample concentration from the designated standard curve, correcting each concentration for associated blank response and also for any user or instrument specified dilution. Dilution by the instrument is noted by software as analysis ensues and, also, documented in the Excel data report file. The analyst examines each row of data. Results are eliminated that are outside the limits of the calibration range, or have an unrepeatable blank response measurement greater than 0.005 absorbance units.
- 12.2 The analyst examines all data that were diluted and reanalyzed to obtain a diluted sample salinity of ≤ 10 ppt. If the response of the diluted sample is > 0.050 absorbance units and the appropriate blank response as described in Section 11.10 is also obtained, the results are accepted. If the response of the diluted sample is < 0.050 absorbance units, CBL Nutrient Analytical Services Laboratory's empirically derived salinity correction is applied to the original undiluted reported concentration.
- $$\text{Salinity Corrected mg NH}_4\text{-N/L} = (((100 - (-1.14 \times \text{ppt sample salinity}) + 116)) / 100) + 1) \times \text{uncorrected mg NH}_4\text{-N/L}$$

13 METHOD PERFORMANCE

- 13.1 On 49 separate dates from January through November 2008, Reagent Blanks were performed on NH₄CBL as deionized water analyzed as a sample. This produced a mean value of 0.0032 mg NH₄-N/L, SD 0.0048.
- 13.2 On 16 separate dates from November 2008 through February 2009, 15 replicate analyses of SPEX® Corporation QC 6-50 NUT 1 were performed on NH₄CBL2. This produced a mean value of 0.2078 mg NH₄-N/L, SD 0.0253, Relative Percent Difference of 4% from the expected value of $0.200 \pm 10\%$. This is a mean recovery of 104%.
- 13.3 For some estuarine samples analyzed on NH₄CBL in 2008, the mean difference in concentration between 76 duplicates analyzed on 13 separate dates was 0.0027 mg NH₄-N/L. The standard deviation of the difference between duplicates was 0.0030 NH₄-N/L.

14 REFERENCES

- 14.1 USEPA. 1979. Method No. 353.2 *in* Methods for chemical analysis of water and wastes. United States Environmental Protection Agency, Office of Research and Development. Cincinnati, Ohio. Report No. EPA-600/4-79-020 March 1979. 460pp.
- 14.2 Frank, J. M., C.F. Zimmermann and C. W. Keefe (2006). Comparison of results from Konelab Aquakem 250 and existing nutrient analyzers. UMCES CBL Nutrient Analytical Services Laboratory, Dec. 2006.
- 14.3 Solorzano, L. 1969. Determination of ammonia in natural waters by the phenolhypochlorite method. *Limnol. Oceanogr.* 14:799-801.
- 14.4 Parsons, T.R., Y. Maita and C.M. 1984. *A Manual of Chemical and Biological Methods for Seawater Analysis*, Pergamon Press, Elmsford, N.Y.
- 14.5 Kerouel, R. and A. Aminot. 1987. Procédure optimisée hors-contaminations pour l'analyse des éléments nutritifs dissous dans l'eau de mer. *Mar. Environ. Res* 22:19-32.